



Genomic hotspots of adaptation in butterfly wing pattern evolution

Riccardo Papa, Arnaud Martin and Robert D Reed

What is the genetic architecture of morphological evolution? Is there uniform potential for novelty across a genome or, on the contrary, can a small number of large-effect genes explain the phenotypic variation observed within and between species? Here we highlight recent work on butterfly wing pattern genetics showing that a small set of loci can be repeatedly involved in the evolution of complex traits. These loci behave as genomic hotspots for diversification because they underlie adaptive variation within and between species with both convergent and highly divergent wing patterns. These findings suggest that certain loci may be more likely than others to facilitate rapid evolutionary change.

Address

Department of Ecology and Evolutionary Biology, University of California, Irvine, 321 Steinhaus Hall, Irvine, CA 92697-2525, USA

Corresponding author: Reed, Robert D (rreed@uci.edu)

Current Opinion in Genetics & Development 2008, 18:559-564

This review comes from a themed issue on Genomes and evolution Edited by Sarah Teichmann and Nipam Patel

Available online 8th January 2009

0959-437X/\$ - see front matter
© 2008 Elsevier Ltd. All rights reserved.

DOI 10.1016/j.gde.2008.11.007

Introduction

The last few years have seen remarkable advances in understanding the molecular basis of morphological evolution. Widespread access to new technologies has made it possible to identify the genomic targets of selection that control adaptive phenotypes, including in species not normally considered to be genetic model systems. This has now set the stage to ask more general questions about the genetic architecture of evolving phenotypes [1,2]. One of these questions is whether certain genes tend to underlie phenotypic adaptation more than others. Are there certain loci — genomic hotspots of adaptation — that make a disproportionate contribution to evolutionary change across a phylogenetic spectrum?

The recent literature regarding the genetic basis of convergent and parallel evolution in animals is of particular interest because it reveals some potential examples of adaptive hotspots — instances where the same gene

underlies adaptation in multiple lineages. There are now numerous cases known of specific genes underlying parallel and convergent evolution [3]. For example, regulatory elements of yellow underlie multiple cases of convergent evolution of both abdomen and wing pigmentation in *Drosophila* species [4,5], independent events of albinism in cavefish were linked to mutations in Oca2, a determinant of pigmentation in human populations [6,7], and derived pigmentation in several stickleback populations is linked to a regulatory allele of *Kitlg*, a gene associated with skin color in humans [8]. Repeated fixation of an allele of the Eda gene is responsible for the reduction of armor plates in sticklebacks [9-11], while its receptor Edar matches a quantitative trait locus (QTL) for hair thickness in humans [12]. Modulation of Pitx1 expression explains repeated pelvic reduction in two different species of sticklebacks, and is also thought to be responsible for pelvic reduction in manatee, a marine mammal [13,14]. Also, Bmp4 has been proposed as a hotspot for the evolution of feeding strategies since it acts as a QTL influencing the mechanical properties of cichlid fish mandibles [15] and presumably the beaks of Darwin's finches [16]. These are all examples of loci repeatedly exploited by evolution to produce similar adaptive traits in different lineages.

In retrospect it is not surprising that the evolution of similar phenotypes often occurs through changes in the same genes. At this point there are enough examples of specific genes underlying parallel and convergent evolution that it might appear to be a general evolutionary trend. It has been noted, however, that there are a similar number of examples of convergence occurring through changes in different genes [3]. Accordingly, to identify potential adaptive hotspots it may be more useful to ask if some loci tend to underlie the evolution of different morphologies across multiple lineages. This is a phenomenon for which there are fewer examples. One case of this is the Mc1r gene, in which coding mutations trigger divergent patterns of melanization in mammals, birds, and lizards, including at the population level [17,18]. Another possible example is that regulatory evolution of svb/ovo explains multiple cases of convergence in the larval bristle patterns of species of *Drosophila* [19,20], while the nematode ortholog of svb/ovo underlies the evolution of excretory duct morphology in Caenorhabditis elegans [21]. In this latter example, however, it is questionable whether comparing only two taxa that are so phylogenetically distant represents a trend rather than a coincidence.

Despite the case studies described above, one can argue that the adaptive hotspot hypothesis is premature, or the result of a 'street light syndrome', where inferences are biased by expectations. This is especially a concern in studies relying on a candidate gene approach (i.e. focusing on gene in one system because its function is known in another system), which many of the above examples do. In this respect, we argue that butterfly wing patterns are a uniquely powerful system for assessing potential adaptive hotspots. Genetic mapping efforts are underway in several different butterfly species [22,23], allowing completely independent discoveries of loci controlling wing pattern variation and divergence in nature. This approach precludes biases resulting from candidate genes. Furthermore, the links between morphological variation and adaptive evolution are particularly well known in butterflies. Indeed, butterfly wing pattern genes are frequently known to be under selection because they play a clear role in adaptation — particularly in situations involving mimicry, crypsis, and mate choice. For these reasons, loci controlling natural variation in wing patterns are exceptionally useful as 'model genes' for comparative studies of adaptation.

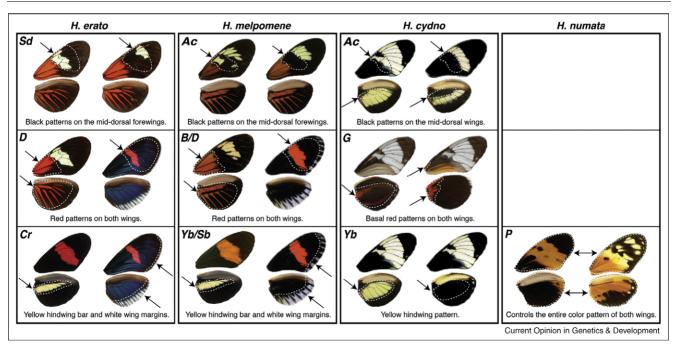
Color pattern genetics of Heliconius butterflies

While color pattern related linkage mapping is being done for several butterfly species, most published work

is from the neotropical nymphalid butterfly genus Heliconius — the group we focus on here. The most recent phylogenetic work places 38 species in this genus [24], most of which are highly polymorphic and participate in mimicry [25]. *Heliconius* is most famous for the spectacular convergent radiations between the two distantly related comimics Heliconius erato and Heliconius melpomene [26,27]. In this textbook example of Müllerian mimicry (i.e. mimicry where all participants are distasteful) each species consists of at least 20 named geographic races, most of which mimic co-occurring races of the other Heliconius species, as well as other species of butterflies and day-flying moths. Population genetic work suggests that H. erato represents an older radiation and that *H. melpomene* evolved to resemble *H. erato*'s wing patterns [28].

Over the last 50 years intraspecific crossing experiments using different wing pattern races of *H. erato* and *H.* melpomene have led to the description of a toolkit of more than a dozen Mendelian genes controlling natural wing pattern variation in each species [29°,30°,31°,32–35]. Even though many genes affecting color patterns have been named, much of the color pattern variation in each species can be attributed to only a few loci of major effect (Figure 1). Efforts to map these color patterns genes have focused on four species: H. erato, H. melpomene, Heliconius

Figure 1



A brief outline of the effects of three major color pattern loci across four different Heliconius species. Each column represents a different species, while each row represents a homologous color pattern locus. Names of loci are in upper left corners of panels, and arrows and dashed lines specify the regions of effect. All wings are presented by their dorsal surfaces with the exception of the H. cydno G locus panel, for which the ventral surface is shown. Note that in H. melpomene B and D are tightly linked loci, where B controls the color of the forewing band, and D controls the hindwing rays and red at the base of the forewing. Similarly, in H. melpomene Yb and Sb are tightly linked loci, where Yb controls the hindwing yellow bar, and Sb controls the white wing margins. In H. numata the entire color pattern variation is controlled by a single Mendelian locus, P.

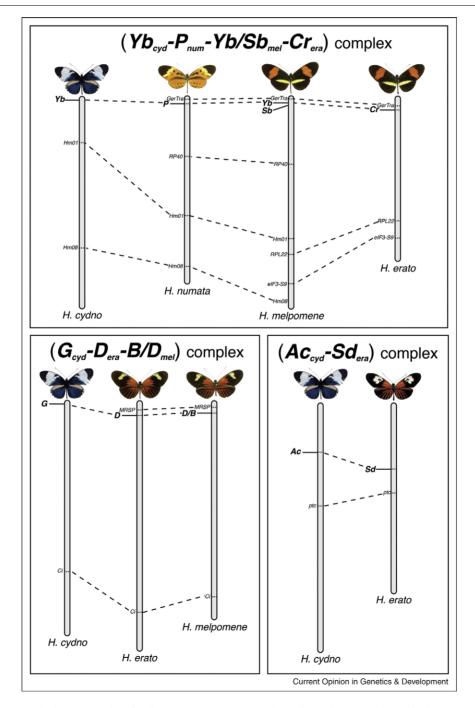
cydno, and Heliconius numata. Of these, the H. erato lineage is distantly related to the lineage containing the three closely related species H. melpomene, H. cydno, and H. numata. This sampling of species represents an intentional strategy to compare distantly related species with convergent phenotypes (H. erato and H. melpomene), as well as closely related species with highly divergent

phenotypes (H. melpomene, H. cydno, and H. numata) [27,36°,37°].

Homologous loci control convergent and divergent wing patterns across species

The most remarkable finding to emerge from mapping efforts in Heliconius is that the same few loci appear to

Figure 2



Comparative linkage mapping in different species of Heliconius suggests that a small set of homologous loci control both convergent and divergent color patterns across species. The Yb-P-Yb/Sb-Cr locus controls multiple pattern elements in a complex way across four species, the G-D-B/D locus controls various basal red patterns in at least three species, and the Ac-Sd locus controls forewing melanin patterns in a complex way in at least two species.

underlie the majority of natural variation across all of the species studied so far (Figure 2). This is perhaps best exemplified by Joron et al. who showed that the Cr locus of H. erato maps to the same location as Yb/Sb in H. melpomene [36**]. This in itself may not be completely surprising because these loci control similar color pattern elements in the two species, and there is a strong precedent for a conserved genetic basis of convergent evolution as we previously discussed. The most interesting discovery, however, was that the P locus, which controls wing pattern variation in *H. numata*, mapped to the very same position [36**]. H. numata is closely related to H. melpomene, but it looks completely different and its entire wing pattern is controlled by a single locus with many major-effect alleles, as opposed to a multi-gene toolkit as in H. melpomene.

This phenomenon extends to other loci and species as well. Kronforst et al. mapped three color pattern genes in *H. cydno*: Yb, Ac, and G [37 $^{\bullet \bullet}$]. H. cydno is closely related to H. melpomene and H. numata, but has very different wing patterns. Yb in H. cydno has a similar effect to Yb in H. melpomene — it controls the presence and absence of a yellow hindwing bar. It was therefore unsurprising to find that Yb maps to the same locus in both species. More surprising was that Ac in H. cydno maps to the H. erato Sd locus. The effect of this locus is fairly different in the two species, except in both species it is involved in controlling the melanic pattern elements on the forewing. Another surprise was that the G locus maps to H. erato's D, and by extension, H. melpomene's B/D. G phenotypes in H. cydno do not closely resemble D phenotypes, except that they both influence red patterns in the basal regions of wings.

These comparative mapping data are extremely interesting; however, the specific molecular identities of the Heliconius color pattern genes remain unknown. Accordingly, research in several labs is now focused on positionally cloning these loci. To date, researchers have characterized the ~300 kb sequence interval including Cr-Yb and the ~ 600 kb interval including D-B/D in both H. erato and H. melpomene [38°,39°°]. Comparative sequence analysis of these regions shows almost perfect synteny in predicted genes between H. erato, H. melpomene, and the silkmoth Bombyx mori. This finding not only suggests that synteny is highly conserved across the Macrolepidoptera, but it also supports the hypothesis that the *Heliconius* color pattern loci are homologous between species at the DNA sequence level. The next challenge is to identify the specific recombining sequences underlying *Heliconius* color pattern variation. This will not only be informative as to what kind of genetic elements adaptive hotspots may tend to be (e.g. protein-coding regions, cis-regulatory regions, or regulatory RNAs), but it will also allow a better understanding regarding the age and diversity of the variationcausing alleles relative to the color pattern radiations themselves.

Portrait of a genomic hotspot for morphological evolution

Presumably it takes dozens or hundreds of genes to make a butterfly wing pattern. If this is so, why do such a small number of genes repeatedly underlie the evolution of wing patterns? One possible explanation is that developmental and genetic constraints channel evolutionary change in such a way that only a small number of loci have significant potential to produce novel phenotypes. One situation that would produce this effect would be if adaptive hotspots tended to be minimally pleiotropic downstream effector genes. Functional changes in structural genes could be relatively difficult to buffer against developmentally and would often have large effects in specific tissues. There is a strong precedent for this model in other proposed hotspots: Mc1r and Oca2 are specific to the pigmentation process [18], and Edar is restricted to skin appendage development [40]. This model probably does not apply to *Heliconius*, however. Although regulation of the pigment gene vermilion appears to be controlled by D in H. erato [41], thus far there are no obvious pigmentation genes in the color pattern gene sequence intervals [38°,39°°] or yet identified as being genetically linked to the color pattern loci [29°,30°,31°].

Another scenario that would focus evolutionary change on a few loci would be if key mutations occurred in highly pleiotropic genes that act through controlling the expression of downstream developmental modules. Largeeffect tissue-specific changes could be achieved through changes in the regulatory regions of such upstream regulatory genes [4,19,21,42,43]. Indeed, modeling work has suggested that regulatory networks should tend to evolve an architecture where the majority of the network is robust to mutation, but mutations at specific hub genes in the network will have large effects tending to be relatively benign [44]. Work on the Edar network in sticklebacks has also led to the proposal that these hub genes should have larger and more modular cis-regulatory regions, and thus would be more likely to undergo regulatory evolution [45]. On the basis of these ideas one might predict that the Heliconius color pattern loci represent relatively upstream developmental regulatory genes with complex *cis*-regulatory regions.

Conclusion

Mapping and positional cloning work across the genus Heliconius has shown that a small set of homologous loci repeatedly underlie the evolution of both convergent and divergent wing patterns within and between species. It remains to be seen, however, exactly what the molecular elements underlying this variation are. Further fine-scale mapping, nucleotide polymorphism association studies, comparative sequencing, and gene expression studies will hopefully allow us to better understand the nature of these adaptive hotspots in the butterfly genome. Ongoing work will be greatly facilitated by the continuing development of genomic resources like BAC libraries, EST databases, and expression microarrays [23], including the scheduled sequencing of two *Heliconius* genomes in 2009.

Future work on other butterfly species will give us a notion of how 'hot' these genomic hotspots of adaptation really are. Do these hotspots extend into other genera or families of butterflies? Many species of butterflies are polymorphic — is it possible that these *Heliconius* hotspots also underlie variation in other groups of butterflies? Comparative analysis of *Heliconius* color pattern polymorphism, modular control of eyespot size in Bicyclus butterflies [46,47], and swallowtail mimicry [48] could allow the first steps into gaining insight into this broader question. Indeed, the genetic mechanisms that lead to wing pattern diversity could be more directly reflective of an evolutionary ground plan [49] than previously thought.

Acknowledgements

We thank Owen McMillan for comments on the manuscript, and Marcus Kronforst and Chris Jiggins for helpful discussions.

References and recommended reading

Papers of particular interest, published within the period of the review, have been highlighted as:

- · of special interest
- of outstanding interest
- Hoekstra HE, Coyne JA: The locus of evolution: evo devo and the genetics of adaptation. Evolution 2007, 61:995-1016.
- Wray GA: The evolutionary significance of cis-regulatory mutations, Nat Rev Genet 2007, 8:206-216.
- Arendt J, Reznick D: Convergence and parallelism reconsidered: what have we learned about the genetics of adaptation? Trends Ecol Evol 2008, 23:26-32.
- Jeong S, Rokas A, Carroll SB: Regulation of body pigmentation by the abdominal-B Hox protein and its gain and loss in Drosophila evolution. Cell 2006, 125:1387-1399.
- Prud'homme B, Gompel N, Rokas A, Kassner VA, Williams TM, Yeh SD, True JR, Carroll SB: Repeated morphological evolution through cis-regulatory changes in a pleiotropic gene. Nature 2006, 440:1050-1053.
- Norton HL, Kittles RA, Parra E, McKeigue P, Mao XY, Cheng K, Canfield VA, Bradley DG, McEvoy B, Shriver MD: **Genetic** evidence for the convergent evolution of light skin in Europeans and east Asians. Mol Biol Evol 2007, **24**:710-722
- Protas ME, Hersey C, Kochanek D, Zhou Y, Wilkens H, Jeffery WR, Zon Ll, Borowsky R, Tabin CJ: Genetic analysis of cavefish reveals molecular convergence in the evolution of albinism. Nat Genet 2006, 38:107-111.
- Miller CT, Beleza S, Pollen AA, Schluter D, Kittles RA, Shriver MD, Kingsley DM: cis-regulatory changes in kit ligand expression and parallel evolution of pigmentation in sticklebacks and humans. Cell 2007, 131:1179-1189.
- Colosimo PF, Hosemann KE, Balabhadra S, Villarreal G, Dickson M, Grimwood J, Schmutz J, Myers RM, Schluter D, Kingsley DM: Widespread parallel evolution in sticklebacks by repeated fixation of ectodysplasin alleles. Science 2005, **307**:1928-1933.
- 10. Kitano J, Bolnick DI, Beauchamp DA, Mazur MM, Mori S, Nakano T, Peichel CL: Reverse evolution of armor plates in the threespine stickleback. Curr Biol 2008, 18:769-774.

- 11. Cresko WA, Amores A, Wilson C, Murphy J, Currey M, Phillips P, Bell MA, Kimmel CB, Postlethwait JH: Parallel genetic basis for repeated evolution of armor loss in Alaskan threespine stickleback populations. Proc Natl Acad Sci U S A 2004, 101:6050-6055
- 12. Fujimoto A, Kimura R, Ohashi J, Omi K, Yuliwulandari R, Batubara L, Mustofa MS, Samakkarn U, Settheetham-Ishida W, Ishida T et al.: A scan for genetic determinants of human hair morphology: EDAR is associated with Asian hair thickness. Hum Mol Genet 2008, 17:835-843.
- Shapiro MD, Bell MA, Kingsley DM: Parallel genetic origins of pelvic reduction in vertebrates. Proc Natl Acad Sci U S A 2006, . **103**:13753-13758.
- 14. Shapiro MD, Marks ME, Peichel CL, Blackman BK, Nereng KS, Jonsson B, Schluter D, Kingsley DM: Genetic and developmental basis of evolutionary pelvic reduction in threespine sticklebacks. Nature 2004, 428:717-723.
- 15. Albertson RC, Streelman JT, Kocher TD, Yelick PC: Integration and evolution of the cichlid mandible: the molecular basis of alternate feeding strategies. Proc Natl Acad Sci U S A 2005, 102:16287-16292
- 16. Abzhanov A, Protas M, Grant BR, Grant PR, Tabin CJ: Bmp4 and morphological variation of beaks in Darwin's finches. Science 2004. 305:1462-1465.
- 17. Steiner CC, Weber JN, Hoekstra HE: Adaptive variation in beach mice produced by two interacting pigmentation genes. PLoS Biol 2007, 5:1880-1889
- 18. Hoekstra HE: Genetics, development and evolution of adaptive pigmentation in vertebrates. Heredity 2006, 97:222-234
- 19. McGregor AP, Orgogozo V, Delon I, Zanet J, Srinivasan DG, Payre F, Stern DL: Morphological evolution through multiple cis-regulatory mutations at a single gene. Nature 2007, 448:U586-U587.
- 20. Sucena E, Delon I, Jones I, Payre F, Stern DL: Regulatory evolution of shavenbaby/ovo underlies multiple cases of morphological parallelism. Nature 2003, 424:935-938.
- 21. Wang XD, Chamberlin HM: Evolutionary innovation of the excretory system in Caenorhabditis elegans. Nat Genet 2004,
- 22. Marcus JM: Jumping genes and AFLP maps: transforming lepidopteran color pattern genetics. Evol Dev 2005, 7.108-114
- 23. Beldade P, McMillan WO, Papanicolaou A: Butterfly genomics eclosing. Heredity 2008, 100:150-157.
- Beltran M, Jiggins CD, Brower AVZ, Bermingham E, Mallet J: Do pollen feeding, pupal-mating and larval gregariousness have a single origin in Heliconius butterflies? Inferences from multilocus DNA sequence data. Biol J Linn Soc 2007, 92:221-239
- 25. Brown KS: Ecologia Geográfica e Evoluçã nas Florestas Neotropicais. Campinas: Universidade Estadual de Campinas;
- 26. Parchem RJ, Perry MW, Patel NH: Patterns on the insect wing. Curr Opin Genet Dev 2007, 17:300-308.
- 27. Joron M, Jiggins CD, Papanicolaou A, McMillan WO: Heliconius wing patterns: an evo-devo model for understanding phenotypic diversity. Heredity 2006, 97:157-167.
- 28. Flanagan NS, Tobler A, Davison A, Pybus OG, Kapan DD, Planas S. Linares M. Heckel D. McMillan WO: Historical demography of Müllerian mimicry in the neotropical Heliconius butterflies. Proc Natl Acad Sci U S A 2004, **101**:9704-9709.
- Jiggins CD, Mavarez J, Beltran M, McMillan WO, Johnston JS, Bermingham E: A genetic linkage map of the mimetic butterfly Heliconius melpomene. Genetics 2005, 171:557-570.

Placed the Yb and Sb loci on an AFLP-based linkage map of the H. melpomene genome.

30. Kapan DD, Flanagan NS, Tobler A, Papa R, Reed RD, Gonzalez JA, Restrepo MR, Martinez L, Maldonado K, Ritschoff C et al.: Localization of Müllerian mimicry genes on a dense linkage map of Heliconius erato. Genetics 2006, 173:735-757.

Placed the D and Sd color pattern loci on an AFLP-based linkage map of the H. erato genome.

Kronforst MR, Young LG, Kapan DD, McNeely C, O'Neill RJ, Gilbert LE: Linkage of butterfly mate preference and wing color preference cue at the genomic location of wingless. Proc Natl Acad Sci U S A 2006, 103:6575-6580.

This study showed that mate preference maps to the same genomic region as the choice-cue color pattern polymorphism in H. cydno.

- Sheppard PM, Turner JRG, Brown KS, Benson WW, Singer MC: Genetics and the evolution of muellerian mimicry in Heliconius butterflies. Philos Trans R Soc Lond (B) 1985, 308:433-613
- 33. Tobler A, Kapan D, Flanagan NS, Gonzalez C, Peterson E, Jiggins CD, Johntson JS, Heckel DG, McMillan WO: Firstgeneration linkage map of the warningly colored butterfly Heliconius erato. Heredity 2005, 94:408-417.
- 34. Naisbit RE, Jiggins CD, Mallet J: Mimicry: developmental genes that contribute to speciation. Evol Dev 2003, 5:269-280.
- Jiggins CD, McMillan WO: The genetic basis of an adaptive radiation: warning colour in two Heliconius species. Proc R Soc B Biol Sci 1997, 264:1167-1175.
- 36. Joron M, Papa R, Beltran M, Chamberlain N, Mavarez J, Baxter S, Abanto M, Bermingham E, Humphray SJ, Rogers J et al.: A conserved supergene locus controls colour pattern diversity in Heliconius butterflies. PLoS Biol 2006, 4:1831-1840.

This study showed that the Yb, Cr, and P loci map to the same chromosomal positions in H. melpomene, H. erato, and H. numata, respectively. This is particularly important because it demonstrates how a single locus can underlie variation in both convergent and divergent color patterns across multiple species.

37. Kronforst MR. Kapan DD. Gilbert LE: Parallel genetic architecture of parallel adaptive radiations in mimetic Heliconius butterflies. Genetics 2006. 174:535-539.

This comparative mapping study showed that the three H. cydno color pattern genes *G*, *Yb*, and *Ac* map to the same positions as the *H. erato* genes *D*, *Cr*, and *Sd*, respectively. It also showed that *Yb* in *H. cydno* maps to *Yb* in *H. melpomene*. This study is significant because it provides the first comparative mapping work for H. cydno, which is closely related to H. melpomene, but looks very different from the comimics H. melpomene and H. erato.

38. Baxter SW, Papa R, Chamberlain N, Humphray SJ, Joron M, ffrench-Constant RH, McMillan WO, Jiggins CD: Convergent evolution in the genetic basis of Müllerian mimicry in Heliconius butterflies. Genetics 2008, 180:1567-1577.

This study describes the sequence of the B/D color pattern gene interval in H. melpomene, and shows that this region maps to the same region as the D locus in H. erato.

- 39. Papa R, Morrison CM, Walters JR, Counterman BA, Chen R,
 Halder G, Ferguson L, Chamberlain N, ffrench-Constant R,
- Kapan DD et al.: Highly conserved gene order and numerous novel repetitive elements in genomic regions linked to wing pattern variation in Heliconius butterflies. BMC Genomics 2008, **9**:345.

This study describes the genomic sequence intervals of the *D* and *Cr* loci in *H. erato* and shows that these sequences, including gene synteny, are conserved between H. erato, H. melpomene, and B. mori. This is some of the strongest evidence so far that the color pattern genes are homologous at the molecular level.

- 40. Cui CY, Schlessinger D: EDA signaling and skin appendage development. Cell Cycle 2006, 5:2477-2483
- 41. Reed RD, McMillan WO, Nagy LM: Gene expression underlying adaptive variation in *Heliconius* wing patterns: non-modular regulation of overlapping cinnabar and vermilion prepatterns. Proc R Soc B Biol Sci 2008, 275:37-45.
- 42. Jeong S, Rebeiz M, Andolfatto P, Werner T, True J, Carroll SB: The evolution of gene regulation underlies a morphological difference between two Drosophila sister species. Cell 2008, **132**:783-793
- 43. Gompel N, Prud'homme B, Wittkopp PJ, Kassner VA, Carroll SB: Chance caught on the wing: cis-regulatory evolution and the origin of pigment patterns in Drosophila. Nature 2005,
- 44. Crombach A, Hogeweg P: Evolution of evolvability in gene regulatory networks. PLoS Comput Biol 2008, 4:e1000112.
- 45. Knecht AK, Hosemann KE, Kingsley DM: Constraints on utilization of the EDA-signaling pathway in threespine stickleback evolution. *Evol Dev* 2007, 9:141-154.
- 46. Monteiro A, Chen B, Scott LC, Vedder L, Prijs HJ, Belicha-Villanueva A, Brakefield PM: The combined effect of two mutations that alter serially homologous color pattern elements on the fore and hindwings of a butterfly. BMC Genetics 2007. 8:22.
- 47. Monteiro A, Prijs J, Bax M, Hakkaart T, Brakefield PM: Mutants highlight the modular control of butterfly eyespot patterns. Evol Dev 2003, 5:180-187.
- Clark R, Brown SM, Collins SC, Jiggins CD, Heckel DG, Vogler AP: Colour pattern specification in the Mocker swallowtail *Papilio* dardanus: the transcription factor invected is a candidate for the mimicry locus H. Proc R Soc B Biol Sci 2008, **275**:1181-1188
- Nijhout HF: The Development and Evolution of Butterfly Wing Patterns. Washington, DC: Smithsonian Institution Press;